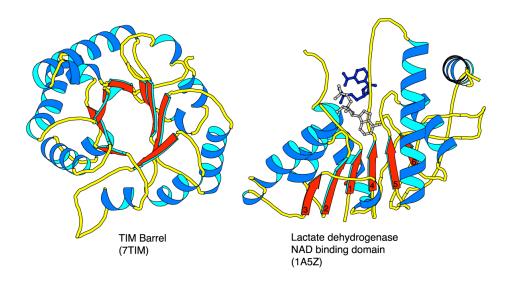
### Protein Structure Determination



How are these structures determined?

## Why Bother With Structure?

- The amino acid sequence of a protein contains interesting information.
- A protein sequence can be compared to other protein sequences to establish its evolutionary relationship to other proteins and protein families.
- However, for the purposes of understanding protein function, the 3D structure of the protein is far more useful than the sequence.

# Protein Sequences Far Outnumber Structures

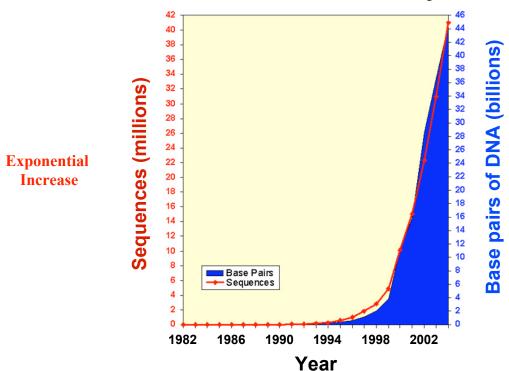
• Only a small number of protein structures have been experimentally determined.

PDB ~30,500 protein structures Genebank ~42,000,000 sequences

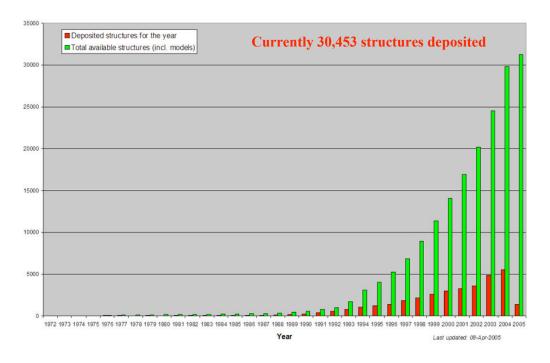
• Of the 30,500 structures, only about **7000** are unique.

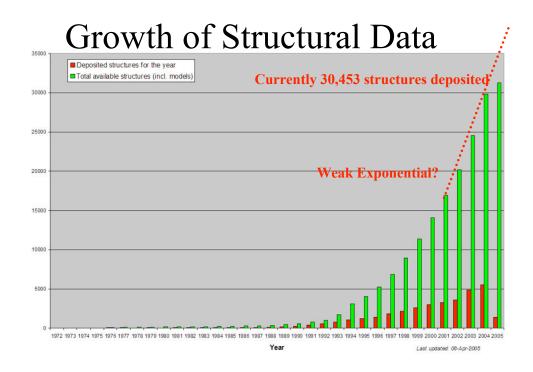
### **Growth of GenBank**

Release 146 (Feb 2005) has 46,849,831,226 base pairs



## Growth of Structural Data





### Structural Proteomics

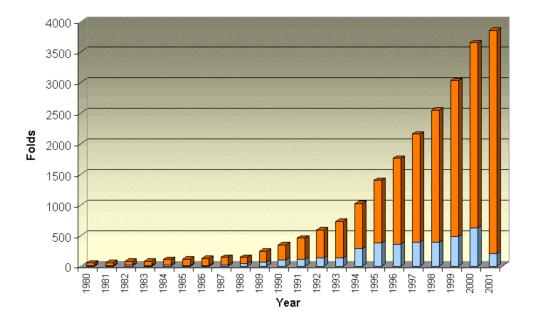
- Use experimentally determined structures to model the structures of similar proteins
  - Threading
    Homology Modeling
    Fold recognition

    Avoids Ab initio structure determination
- Need representative protein structures for the total repertoire of protein folds
- Provide 3D portraits for all proteins in an organism
- Goal: Use structure to infer function.
  - More sensitive than primary sequence comparisons

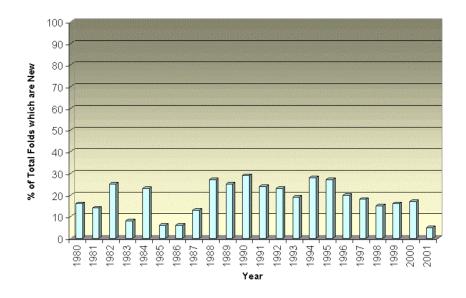
# Redundancy in PDB (19 April 05)

Sequence identity	Number of non-redundant chains				
90%	10503				
70%	9361				
50%	8009				
30%	6120				

# Unique folds in PDB



# New Folds Becoming Rare Why?



### Structural Genomics



Initiated in 1999 by NIH
Phase I included 9 large centers for high
throughput structure determination
Phase I ran from ~2000 - 2005

#### Goal

The long-range goal of the Protein Structure Initiative (PSI) is to make the three-dimensional atomic-level structures of most proteins easily obtainable from knowledge of their corresponding DNA sequences.

http://www.nigms.nih.gov/psi/mission.html

### Structural Genomics

#### **Benefits**

Structural descriptions will help researchers illuminate structure-function relationships and thus formulate better hypotheses and design better experiments.

The PSI collection of structures will serve as the starting point for structure-based drug development by permitting faster identification of lead compounds and their optimization.

The design of better therapeutics will result from comparisons of the structures of proteins that are from pathogenic and host organisms and from normal and diseased human tissues.

The PSI collection of structures will assist biomedical investigators in research studies of key biophysical and biochemical problems, such as protein folding, evolution, structure prediction, and the organization of protein families and folds.

Technical developments, the availability of reagents and materials, and experimental outcome data in protein production and crystallization will directly benefit all structural biologists and provide valuable assistance to a broad range of biomedical researchers.

### Structural Genomics Centers

The Berkeley Structural Genomics Center (BSGC) The BSGC is pursuing an integrated structural genomics program designed to obtain a near-complete structural complement of two minimal genomes, Mycoplasma genitalium and Mycoplasma pneumoniae, two related human and animal pathogens. Both NMR spectroscopy and X-ray crystallography are being used for structural determination.

#### Center for Eukaryotic Structural Genomics (CESG)

The CESG was founded as a collaborative effort to develop the technologies needed for economical high-throughput structure determination of biologically important eukaryotic proteins and to extend the knowledge of fold-function space. This project also aims to further the research of biologically important proteins in Arabidopsis. The protein structures are being determined via X-ray crystallography or NMR spectroscopy.

#### The Joint Center for Structural Genomics (JCSG)

The research focus of the JCSG is on the prokaryote *Thermotoga maritima*, and the eukaryote *Caenorhabditis elegans*, and the main proteins of interest are signaling proteins. The goals involve discovering new protein folds, attaining complete coverage of the proteome of the eubacterium *Thermotoga maritima*, and creating a high-throughput system from the point of target selection through structure determination. X-ray crystallography is being used for structural determination.

#### The Midwest Center for Structural Genomics (MCSG)

The objective of the MCSG is to develop and optimize new, rapid, integrated methods for highly cost-effective determination of protein structures through X-ray crystallography. This project aims to quickly solve a large number of "easy" targets, and in the process develop new, more advanced tools, methods and approaches that can be applied to "unsolved and difficult projects". Protein targets have an emphasis on unknown folds and proteins from disease-causing organisms.

#### e New York Structural Genomics Research Consortium (NYSGRC)

The NYSGRC aims to develop and use the technology for high-throughput structural and functional studies of proteins from humans and model organisms. The consortium is establishing a fully integrated, high-throughput system for protein family classification and target selection, protein expression, purification, crystallization, and structure determination by X-ray crystallography.

### Structural Genomics Centers

#### The Northeast Structural Genomics Consortium (NEGS)

The NEGS is focused on human proteins and proteins from eukaryotic model organisms. The project targets representative proteins to provide "coverage" of fold space, and also proteins that are interesting from a functional genomics perspective. In addition, the center is exploring the complementary aspects of X-ray crystallography and NMR spectroscopy.

#### The Southeast Collaboratory for Structural Genomics (SECSG)

The objective of the SECSG is to develop and test experimental and computational strategies for high throughput structure determination of proteins by X-ray crystallography and NMR methods and to apply these strategies to scan the entire genome of an organism at a rapid pace. The eukaryotic organisms, *Caenorhabditis elegans*, *Homo sapiens* and an ancestrally-related prokaryotic microorganism having a small genome, *Pyrococcus furiosus*, have been selected as representative genomes.

#### Structural Genomics of Pathogenic Protozoa Consortium (SGPP)

The SGPP consortium aims to determine and analyze the structures of a large number of proteins from major global pathogenic protozoa including *Leishmania major*, *Trypanosoma brucei*, *Trypanosoma cruzi* and *Plasmodium falciparum*. These organisms are responsible for the diseases: leishmaniasis, sleeping sickness, Chagas' disease and malaria. X-ray crystallography is being used for structural determination.

#### The TB Structural Genomics Consortium (TB)

The goal of the TB consortium is to determine the structures of over 400 proteins from M. tuberculosis, and to analyze these structures in the context of functional information that currently exists and that is generated by the project. These structures will include about 40 novel folds and 200 new families of protein structures. The protein structures are being determined using X-ray crystallography.

### Protein Structure Databases

Jon

Ingo

Where does protein structural information reside?



http://www.rcsb.org/pdb/

#### - MMDB:

• http://www.ncbi.nlm.nih.gov/Structure/

#### – FSSP:

• http://www.ebi.ac.uk/dali/fssp/

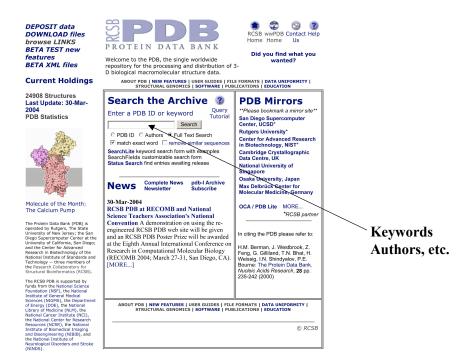
#### **– SCOP**:

• http://scop.mrc-lmb.cam.ac.uk/scop/

#### - CATH:

• http://www.biochem.ucl.ac.uk/bsm/cath\_new/

# http://www.rcsb.org/pdb/

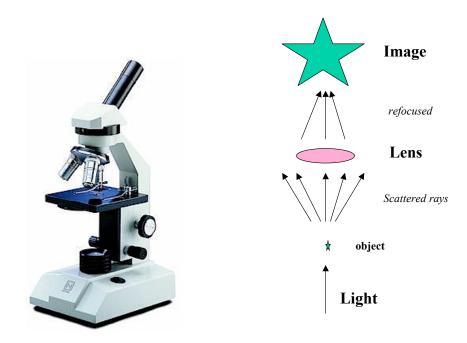


# PDB Contents 19 April 2005

		Molecule Type							
		Proteins, Peptides, and Viruses	Protein/Nu cleic Acid Complexes	Nucleic Acids	Carbohydr ates	Total			
Exp. Tech.	X-ray Diffraction and other	24015	1151	781	11	25958			
	NMR	3733	111	649	2	4495			
	Total	27748	1262	1430	13	30453			

X-ray Crystallography

# Optical Microscope



### **Atomic Resolution**

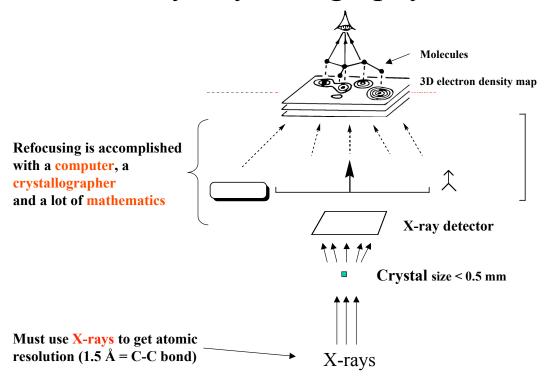
We want to resolve inter-atomic distances (~1.5 Å, 0.15 nM)

Visible light has a wavelength of  $\sim 500$  nm (5000 Å)

Electron beam:  $\lambda_c \sim 0.001~\text{Å}$  (if e<sup>-</sup> is moving at c) Electron velocity is less in electron microscopes Typical resolution is ~10 Å, but can be improved

X-ray generators produce photons of  $\lambda = 0.5 - 2.5 \text{ Å}$ Use  $\lambda = 1.542 \text{ Å}$ 

# X-ray Crystallography



# X-Ray Crystallography

- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

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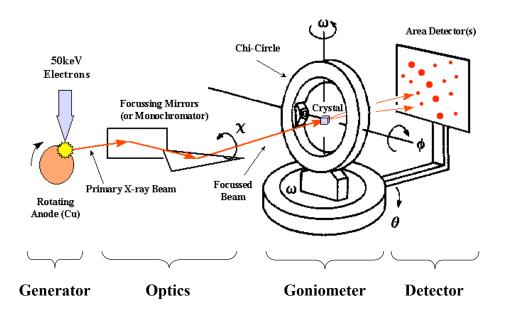
# **Protein Crystals**



# X-Ray Crystallography

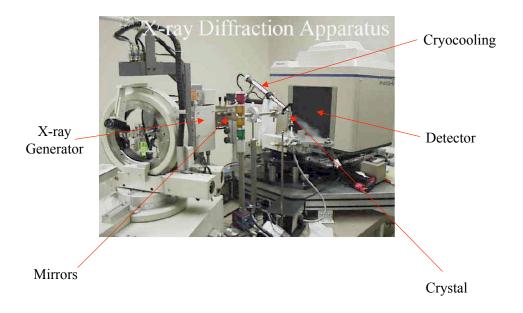
- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

# X-Ray Diffraction Experiment



**Optional: Cryo for protein samples** 

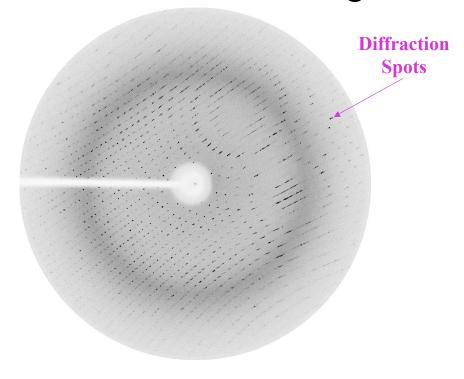
# X-ray Crystallography Equipment



# X-Ray Crystallography

- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

# Protein Diffraction Image



# Why Spots?

X-ray diffraction from individual proteins is diffuse

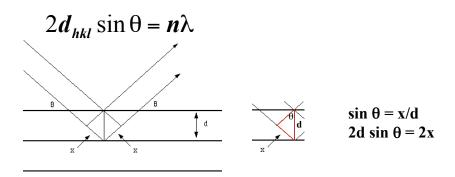
Spots arise due to crystal lattice

**Location** of reflections indicates how an object crystallized 230 possibilities

**Intensity** of reflections contains information about the **structure** of the object in the crystal

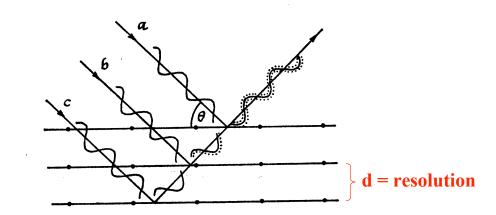
# Bragg's Law

Why do we get spots (reflections) and not a diffuse pattern of scattered x-rays?



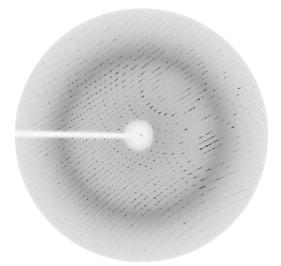
Difference in path (2x) must equal integral number of wavelengths  $(n\lambda)$ 

## Constructive Interference



• Condition for reflection

### Phase Problem



Every diffraction spot (reflection) has a phase and intensity

- -The intensities are recorded by the detector
- -The phases are lost
- -Must have both to reconstruct the image (structure)

### Solutions to the Phase Problem

#### Molecular replacement

- -Use known structure of close homologue
- -Rotational and translational search for solution

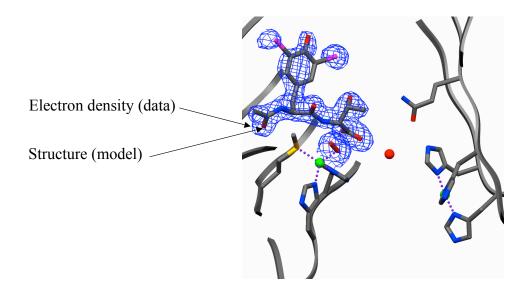
#### Heavy atom labeling

- -Label the protein with electron dense atoms (Hg)
- -Compare independent datasets collected from native and labeled protein
- -Heavy atom substructure provides initial phases

#### **Anomalous diffraction**

- -Crystal must contain atoms with absorption edges between 0.5 and 2.5 Å
- -Compare independent datasets collected at pre-edge and post-edge x-ray energies

# Model Building



# Crystallography Pros/Cons

### **Advantages**

- -can be "fast" down to a few months
- -large structures possible (ribosome)
- -very low resolution (down to 0.5 Å)
- -observables typically > refinement parameters

### **Disadvantages**

- -requires crystal formation
- -non-physiological conditions
- -crystal contacts can limit protein motion

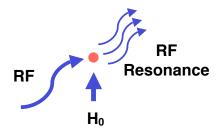
# Nuclear Magnetic Resonance

# Nuclear Magnetic Resonance

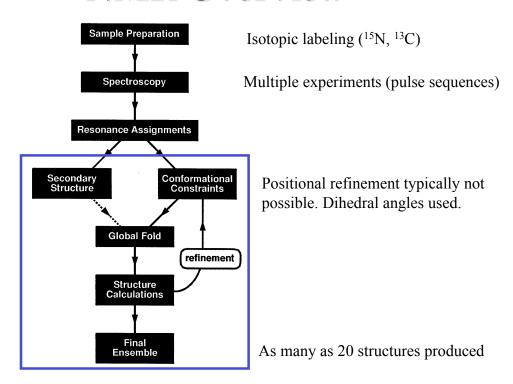
Magnetically align unpaired proton spins (H<sub>0</sub>)

Probe with radio frequency (RF)

#### **Observe resonance**



## **NMR Overview**



# NMR Experimental Observables

- Backbone conformation from chemical shifts (Chemical Shift Index- CSI)
- Distance constraints from NOEs
- Hydrogen bond constraints
- Backbone and side chain dihedral angle constraints from scalar couplings
- Orientation constraints from residual dipolar couplings

### NMR Pros/Cons

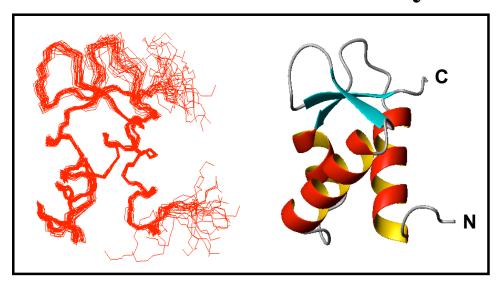
#### **Advantages**

- -no crystal formation needed
- -more physiological conditions

#### **Disadvantages**

- -results in a set of models that are compatible with data
- -size limitation to 200-300 residues (extended recently)
- -must label protein with <sup>15</sup>N and <sup>13</sup>C
- -observables typically < refinement parameters

# Precision NMR vs. X-ray



**RMSD** of the ensemble

Mean coordinate error

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

```
HEADER OXIDOREDUCTASE 03-OCT-02 1MXT

TITLE ATOMIC RESOLUTION STRUCTURE OF CHOLESTEROL OXIDASE

TITLE 2 (STREPTOMYCES SP. SA-COO)

COMPND MOL_ID: 1;

COMPND 2 MOLECULE: CHOLESTEROL OXIDASE;

COMPND 3 CHAIN: A;

COMPND 4 SYNONYM: CHOD;

COMPND 5 EC: 1.1.3.6;

COMPND 6 ENGINEERED: YES;

COMPND 7 OTHER_DETAILS: FAD COFACTOR NON-COVALENTLY BOUND TO THE

COMPND 8 ENZYME
```

### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

```
SOURCE MOL_ID: 1;
SOURCE 2 ORGANISM_SCIENTIFIC: STREPTOMYCES SP.;
SOURCE 3 ORGANISM_COMMON: BACTERIA;
SOURCE 4 GENE: CHOA;
SOURCE 5 EXPRESSION_SYSTEM: ESCHERICHIA COLI;
SOURCE 6 EXPRESSION_SYSTEM_COMMON: BACTERIA;
SOURCE 7 EXPRESSION_SYSTEM_STRAIN: BL21(DE3)PLYSS;
SOURCE 8 EXPRESSION_SYSTEM_VECTOR_TYPE: PLASMID;
SOURCE 9 EXPRESSION_SYSTEM_PLASMID: PCO202
```

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

```
AUTHOR A.VRIELINK,P.I.LARIO
REVDAT 1 25-FEB-03 IMXT 0

JRNL AUTH P.I.LARIO,N.SAMPSON,A.VRIELINK

JRNL TITL SUB-ATOMIC RESOLUTION CRYSTAL STRUCTURE OF

JRNL TITL 2 CHOLESTEROL OXIDASE: WHAT ATOMIC RESOLUTION

JRNL TITL 3 CRYSTALLOGRAPHY REVEALS ABOUT ENZYME MECHANISM AND

JRNL TITL 4 THE ROLE OF FAD COFACTOR IN REDOX ACTIVITY

JRNL REF J.MOL.BIOL. V, 326 1635 2003

JRNL REFN ASTM JMOBAK UK ISSN 0022-2836
```

### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

```
REMARK 3 DATA USED IN REFINEMENT.
REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS): 0.95
REMARK 3 RESOLUTION RANGE LOW (ANGSTROMS): 28.00
REMARK 3 DATA CUTOFF (SIGMA(F)): 0.000
REMARK 3 COMPLETENESS FOR RANGE
                                     (%):94.1
REMARK 3 CROSS-VALIDATION METHOD
                                       : FREE R
REMARK 3 FREE R VALUE TEST SET SELECTION: RANDOM
REMARK 3 FIT TO DATA USED IN REFINEMENT (NO CUTOFF).
REMARK 3 R VALUE (WORKING + TEST SET, NO CUTOFF): 0.110
REMARK 3 R VALUE
                     (WORKING SET, NO CUTOFF): 0.110
REMARK 3 FREER VALUE
                              (NO CUTOFF): 0.132
REMARK 3 FREE R VALUE TEST SET SIZE (%, NO CUTOFF): 5.000
REMARK 3 FREE R VALUE TEST SET COUNT (NO CUTOFF): 13180
REMARK 3 TOTAL NUMBER OF REFLECTIONS (NO CUTOFF): 263551
```

#### **Resolution:**

Low > 3 Å Mid 2-3 Å High 1.5-2 Å Very High < 1.5 Å

#### R factor (residual):

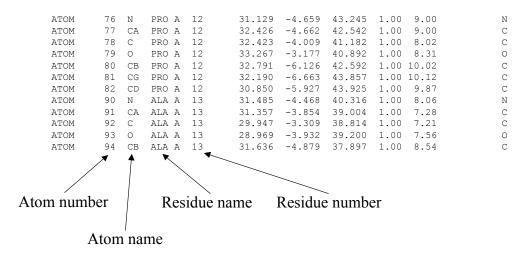
Low resolution  $\sim 27\%$ Mid resolution  $\sim 22\%$ High resolution  $\sim 29\%$ Very High res  $\sim 15\%$ 

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

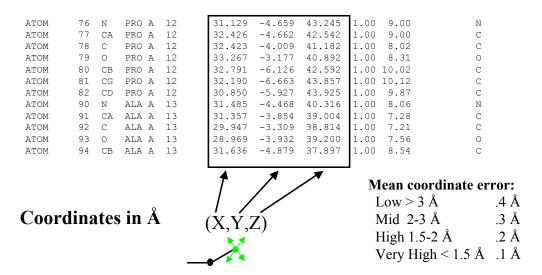
HELIX	14	14 ALA A	289	THR A	304	1					16
HELIX	15	15 THR A	402	GLN A	405	-5					4
HELIX	16	16 ASN A	406	GLY A	425	1					20
HELIX	17	17 ASP A	474	ILE A	478	-5					5
HELIX	18	18 PRO A	486	VAL A	506	1					21
SHEET	1	A 6 HIS	A 248	GLN A	255	0					
SHEET	2	A 6 TYR	A 261	LYS A	268	-1	0	GLU A 266	N	GLN A 249	
SHEET	3	A 6 LEU	A 274	LEU A	287	-1	0	LEU A 275	N	GLN A 267	
SHEET	4	A 6 TYR	A 10	ILE A	16	1	N	VAL A 14	0	PHE A 286	
SHEET	5	A 6 THR	A 36	GLU A	40	1	0	LEU A 37	N	VAL A 15	
SHEET	6	A 6 VAL	A 242	THR A	246	1	0	THR A 243	N	MET A 38	

### A PDB File

Body of PDB file contains information about the atoms in the structure

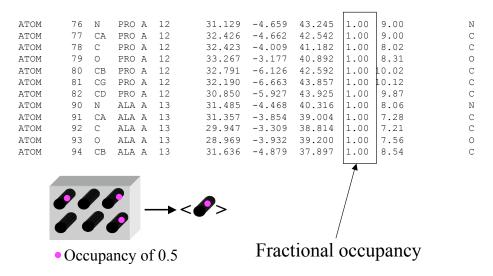


Body of PDB file contains information about the atoms in the structure

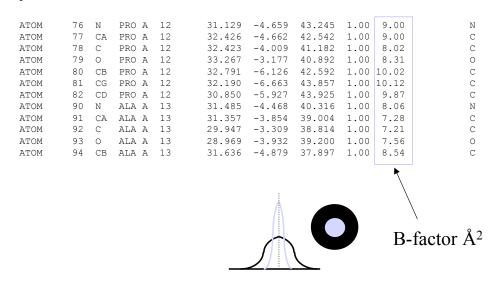


### A PDB File

Body of PDB file contains information about the atoms in the structure



#### Body of PDB file contains information about the atoms in the structure



# Visualization of Structures

