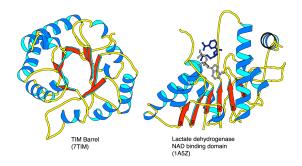
## **Protein Structure Determination**



How are these structures determined?

# Protein Sequences Far Outnumber Structures

• Only a small number of protein structures have been experimentally determined.

PDB ~30,500 protein structures Genebank ~42,000,000 sequences

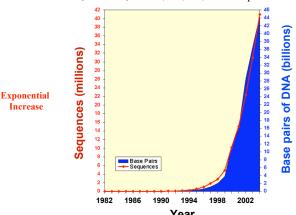
• Of the 30,500 structures, only about **7000** are unique.

# Why Bother With Structure?

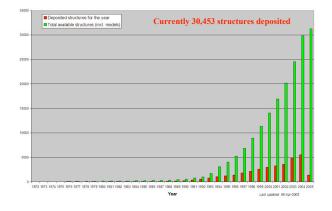
- · The amino acid sequence of a protein contains interesting information.
- A protein sequence can be compared to other protein sequences to establish its evolutionary relationship to other proteins and protein families
- However, for the purposes of understanding protein function, the 3D structure of the protein is far more useful than the sequence.

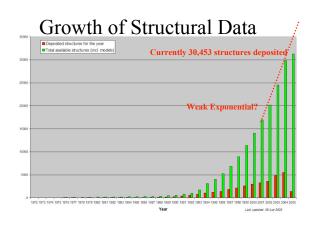
#### Growth of GenBank

Release 146 (Feb 2005) has 46,849,831,226 base pairs



### Growth of Structural Data





#### Structural Proteomics

- Use experimentally determined structures to model the structures of similar proteins
  - ThreadingHomology Modeling

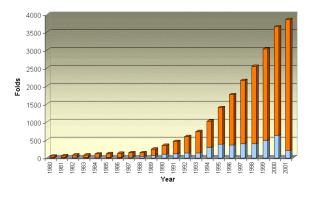
Avoids Ab initio structure determination

- Fold recognition
- Need representative protein structures for the total repertoire of protein folds
- Provide 3D portraits for all proteins in an organism
- Goal: Use structure to infer function.
  - More sensitive than primary sequence comparisons

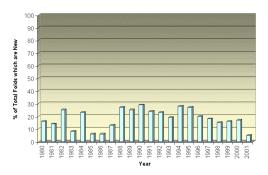
## Redundancy in PDB (19 April 05)

Sequence identity	Number of non- redundant chains
90%	10503
70%	9361
50%	8009
30%	6120

# Unique folds in PDB



# New Folds Becoming Rare Why?



## Structural Genomics



Initiated in 1999 by NIH
Phase I included 9 large centers for high
throughput structure determination
Phase I ran from ~2000 - 2005

#### Goal

The long-range goal of the Protein Structure Initiative (PSI) is to make the three-dimensional atomic-level structures of most proteins easily obtainable from knowledge of their corresponding DNA sequences

http://www.nigms.nih.gov/psi/mission.html

## Structural Genomics

#### Benefit

Structural descriptions will help researchers illuminate structure-function relationships and thus formulate better hypotheses and design better experiments.

The PSI collection of structures will serve as the starting point for structure-based drug development by permitting faster identification of lead compounds and their optimization

The design of better therapeutics will result from comparisons of the structures of proteins that are from pathogenic and host organisms and from normal and diseased human tissues.

The PSI collection of structures will assist biomedical investigators in research studies of key biophysical and biochemical problems, such as protein folding, evolution, structure prediction, and the organization of protein families and folds.

Technical developments, the availability of reagents and materials, and experimental outcome data in protein production and crystallization will directly benefit all structural biologists and provide valuable assistance to a broad range of biomedical researchers.

## Structural Genomics Centers

The Berkels Structural Genomic Center (BSGC). The BSGC is pursuing an integrated structural genomics program designed to obtain a near-complete structural complement of two minimal genomes, Mycoplasma genitalium and Mycoplasma penumoniae, two related human and animal pathogens. Both NMR spectroscopy and X-ray crystallography are being used for structural determination.

The CESG was founded as a collaborative effort to develop the technologies needed for economical high-throughput structure determination of biologically important eukaryotic proteins and to extend the knowledge of fold-function space. This project also aims to further the research of biologically important proteins in Arabidopsis. The protein structures are being determined via X-ray crystallography or NMR spectroscopy.

The research focus of the JCSG is on the prokaryote Thermotoga maritima, and the eukaryote Caenorhabditis elegans, and the main proteins of interest are signaling proteins. The goals involve discovering new protein folds, attaining complete coverage of the proteome of the eubacterium Thermotoga maritima, and creating a high-throughput system from the point of target selection through structure determination. X-ray crystallography is being used for structural determination

The objective of the MCSG is to develop and optimize new, rapid, integrated methods for highly cost-effective determination of protein structures through X-ray crystallography. This project aims to quickly solve a large number of "easy" targets, and in the process develop new, more advanced tools, methods and approaches that can be applied to "unsolved and difficult projects". Protein targets have an emphasis on unknown folds and proteins from disease-causing organisms.

The NYSGRC aims to develop and use the technology for high-throughput structural and functional studies of proteins from humans and model organisms. The consortium is establishing a fully integrated, high-throughput system for protein family classification and target selection, protein expression, purification, crystallization, and structure determination by X-ray crystallography.

#### **Protein Structure Databases**

Where does protein structural information reside?



# PDB Contents 19 April 2005

		Molecule Type						
		Proteins, Peptides, and Viruses	Protein/Nu cleic Acid Complexes	Nucleic Acids	Carbohydr ates	Total		
Exp.	X-ray Diffraction and other	24015	1151	781	11	25958		
	NMR	3733	111	649	2	4495		
	Total	27748	1262	1430	13	30453		

### Structural Genomics Centers

The NEGS is focused on human proteins and proteins from eukaryotic model organisms. The project targets representative proteins to provide "coverage" of fold space, and also proteins that are interesting from a functional genomics perspective. In addition, the center is exploring the complementary aspects of X-ray crystallography and NMR spectroscopy.

The objective of the SECSG is to develop and test experimental and computational strategies for high throughput structure determination of proteins by X-ray crystallography and NMR methods and to apply these strategies to scan the entire genome of an organism at a rapid pace. The eukaryotic organisms, Caenorhabditis elegans, Homo sapiens and an ancestrally-related prokaryotic microorganism having a small genome, Pyrococcus furiosus, have been selected as representative genomes.

The SGPP consortium aims to determine and analyze the structures of a large number of proteins from major global pathogenic protozoa including Leishmania major, Trypanosoma brucei, Trypanosoma cruzi and Plasmodium falciparum. These organisms are responsible for the diseases: leishmaniasis, sleeping sickness, Chagas' disease and malaria. X-ray crystallography is being used for structural determination.

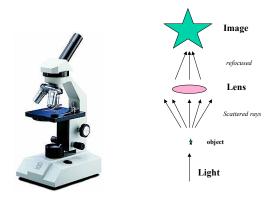
The goal of the TB consortium is to determine the structures of over 400 proteins from M. tuberculosis, and to analyze these structures in the context of functional information that currently exists and that is generated by the project. These structures will include about 40 novel folds and 200 new families of protein structures. The protein structures are being determined using X-ray crystallography.

## http://www.rcsb.org/pdb/

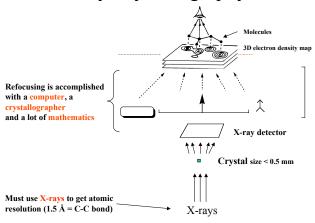


X-ray Crystallography

# Optical Microscope



# X-ray Crystallography



# X-Ray Crystallography

- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

## **Atomic Resolution**

We want to resolve inter-atomic distances (~1.5 Å, 0.15 nM)

Visible light has a wavelength of  $\sim 500$  nm (5000 Å)

Electron beam:  $\lambda_c \sim 0.001$  Å (if e<sup>-</sup> is moving at c) Electron velocity is less in electron microscopes Typical resolution is ~10 Å, but can be improved

X-ray generators produce photons of  $\lambda = 0.5 - 2.5$  Å Use  $\lambda = 1.542$  Å

## X-Ray Crystallography

- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

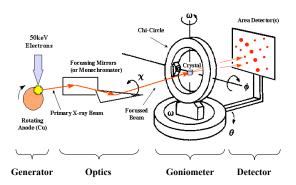
# **Protein Crystals**



## X-Ray Crystallography

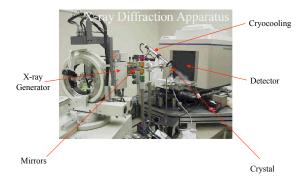
- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

# X-Ray Diffraction Experiment



Optional: Cryo for protein samples

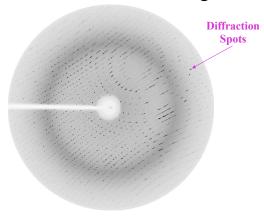
# X-ray Crystallography Equipment



## X-Ray Crystallography

- Make crystals of your protein
   0.3-1.0mm in size
   Proteins must be in an ordered, repeating pattern.
- 3. X-ray beam is aimed at crystal and data is collected.
- 4. Structure is determined from the diffraction data.

# Protein Diffraction Image



# Why Spots?

X-ray diffraction from individual proteins is diffuse

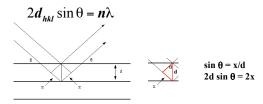
Spots arise due to crystal lattice

Location of reflections indicates how an object crystallized 230 possibilities

Intensity of reflections contains information about the structure of the object in the crystal

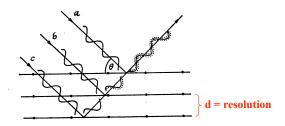
# Bragg's Law

Why do we get spots (reflections) and not a diffuse pattern of scattered x-rays?



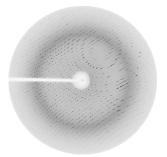
Difference in path (2x) must equal integral number of wavelengths ( $n\lambda$ )

## Constructive Interference



· Condition for reflection

## Phase Problem



Every diffraction spot (reflection) has a phase and intensity

- -The intensities are recorded by the detector
- $\hbox{-The phases are lost}\\$
- -Must have both to reconstruct the image (structure)

## Solutions to the Phase Problem

#### Molecular replacement

- -Use known structure of close homologue
- -Rotational and translational search for solution

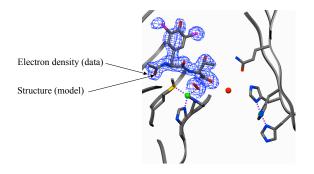
#### Heavy atom labeling

- -Label the protein with electron dense atoms (Hg)
- -Compare independent datasets collected from native and labeled protein
- -Heavy atom substructure provides initial phases

#### **Anomalous diffraction**

- -Crystal must contain atoms with absorption edges between 0.5 and 2.5 Å
- -Compare independent datasets collected at pre-edge and post-edge x-ray energies

# Model Building



# Crystallography Pros/Cons

#### Advantages

- -can be "fast" down to a few months
- -large structures possible (ribosome)
- -very low resolution (down to 0.5 Å)
- -observables typically > refinement parameters

#### Disadvantages

- -requires crystal formation
- -non-physiological conditions
- -crystal contacts can limit protein motion

# Nuclear Magnetic Resonance

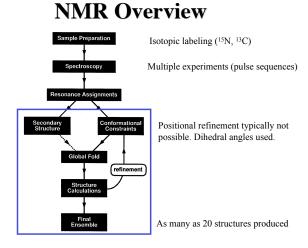
Magnetically align unpaired proton spins (H<sub>0</sub>)

Probe with radio frequency (RF)

Observe resonance



Nuclear Magnetic Resonance



# NMR Experimental Observables

- Backbone conformation from chemical shifts (Chemical Shift Index- CSI)
- · Distance constraints from NOEs
- · Hydrogen bond constraints
- Backbone and side chain dihedral angle constraints from scalar couplings
- Orientation constraints from residual dipolar couplings

# NMR Pros/Cons

#### **Advantages**

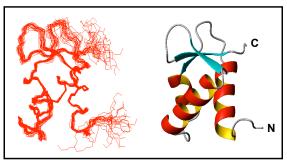
-no crystal formation needed -more physiological conditions

#### **Disadvantages**

- -results in a set of models that are compatible with data
- -size limitation to 200-300 residues (extended recently)
- -must label protein with  $^{15}\mbox{N}$  and  $^{13}\mbox{C}$
- -observables typically < refinement parameters

# Precision

NMR vs. X-ray



RMSD of the ensemble

Mean coordinate error

#### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

HEADER OXIDOREDUCTASE 03-OCT-02 IMXT TITLE ATOMIC RESOLUTION STRUCTURE OF CHOLESTEROL OXIDASE TITLE 2 (STREPTOMYCES SP. SA-COO) COMPND. MOL\_HB-1; COMPND 2 MOLECULE: CHOLESTEROL OXIDASE;
COMPND 3 CHAIN: A, COMPND 4 SYNONYM: CHOD: COMPNO 5 EC: 1.1.3.6; COMPNO 5 EC: 1.1.3.6; COMPNO 6 ENGINEERED: YES; COMPNO 7 OTHER, DETAILS: FAD COFACTOR NON-COVALENTLY BOUND TO THE COMPND 8 ENZYME

#### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

SOURCE MOL\_ID: 1; SOURCE 2 ORGANISM\_SCIENTIFIC: STREPTOMYCES SP.; SOURCE 3 ORGANISM\_COMMON: BACTERIA; SOURCE 4 GENE:-CHOA; SOURCE 4 GENE CHOA.
SOURCE 5 EXPRESSION SYSTEM ESCHERICHIA COLL
SOURCE 6 EXPRESSION SYSTEM COMMON BACTERIA,
SOURCE 7 EXPRESSION SYSTEM STRAIN: BL2(DE3)PLYSS,
SOURCE 8 EXPRESSION SYSTEM VECTOR, TYPE: PLASMID,
SOURCE 9 EXPRESSION SYSTEM PLASMID: PCO202

#### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure eleme

AUTHOR A.VRIELINK,P.I.LARIO REVDAT 1 25-FEB-03 IMXT 0 IOR A VRILLINK, P.I.LARIO
AT 1.2-FEB.20 JIMXT 0
AUTH P.I.LARION, SAMPSON, A VRIELINK
TITL. SUB-ATTOMIC RESOLUTION CRYSTAL STRUCTURE OF
TITL. 2 CHOLESTEROL OXIDASE. WHAT A TOMIC RESOLUTION
TITL. 3 CRYSTALLOGRAPH REVEALS ABOUT FRAYTHE MECHANISM AND
TITL. 4-THE ROLE OF FAD COFACTOR IN REDOX ACTIVITY
REF J.MOL. BIOL. V. 326 1635 2003
REFN ASTM JMOBAK UK ISSN 0022-2836

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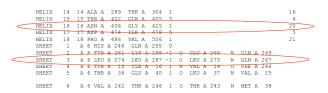
Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements

Resolution: REMARK 3 DATA LISED IN REFINEMENT
FEMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS): 0.95
REMARK 3 RESOLUTION RANGE HIGH (ANGSTROMS): 28.00
REMARK 3 RESOLUTION RANGE LOW (ANGSTROMS): 28.00
REMARK 3 DATA CHIPPE (SIGMAPP): 0.000
REMARK 3 COMPLETENESS FOR RANGE (%): 94.1
REMARK 3 CORSE-VALIDATION METHOD : FREE R
REMARK 3 FREE R VALUE TEST SET SELECTION : RANDOM
REMARK 3 FREE R VALUE FIRST SET SELECTION : RANDOM
REMARK 3 RALLIE (WORKING + TEST SET, NO CUTOFF): 0.110
REMARK 3 R VALUE (WORKING + TEST SET, NO CUTOFF): 0.110
REMARK 3 REEE R VALUE TEST SET SIZE (%, NO CUTOFF): 5.02
REMARK 3 FREE R VALUE TEST SET SIZE (%, NO CUTOFF): 5.130 REMARK 3 DATA USED IN REFINEMENT. REMARK 3 FREE R VALUE TEST SET COUNT (NO CUTOFF): 13180 High resolution ~ 29 % REMARK 3 TOTAL NUMBER OF REFLECTIONS (NO CUTOFF): 263551 Very High res ~ 15%

Low > 3 ÅMid 2-3 Å High 1.5-2 Å Very High < 1.5 Å R factor (residual): Low resolution ~ 27% Mid resolution ~ 22 %

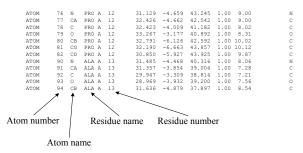
#### A PDB File

Header contains information about protein and structure date of the entry, references, crystallographic data, contents and positions of secondary structure elements



#### A PDB File

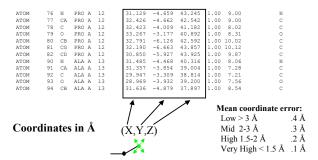
Body of PDB file contains information about the atoms in the structure



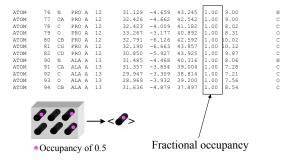
### A PDB File

### A PDB File

#### Body of PDB file contains information about the atoms in the structure

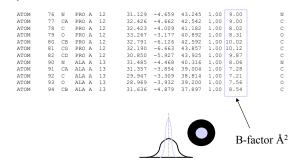


#### Body of PDB file contains information about the atoms in the structure



### A PDB File

#### Body of PDB file contains information about the atoms in the structure



## **Visualization of Structures**

